## Prearranged Glycosides, 9[+]

# Chemical Synthesis of a Tetrasaccharide Fragment Related to the Exopolysaccharide of *Arthrobacter sp.* CE-17

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The tetrasaccharide 5-aminopentyl glycoside  $\beta$ -D-Manp- $(1\rightarrow 4)$ - $\beta$ -D-Glcp- $(1\rightarrow 4)$ - $\alpha$ -L-Rhap- $(1\rightarrow 3)$ - $\beta$ -D-Glcp-1-O- $(CH_2)_5NH_2(\mathbf{22})$  related to the exopolysaccharide of Arthrobacter sp. CE-17 was synthesized by coupling of the properly protected disaccharide blocks  $\beta$ -D-Manp- $(1\rightarrow 4)$ - $\beta$ -D-Glcp-1-S-Ph  $(\mathbf{11})$  and  $\alpha$ -L-Rhap- $(1\rightarrow 3)$ - $\beta$ -D-Glcp-1-O- $(CH_2)_5NHZ <math>(\mathbf{20})$ . Building block  $\mathbf{11}$  was obtained by intramolecular  $\beta$ -mannosylation of a malonyl-tethered disaccharide glycoside which was prepared from phenyl 4,6-

O-benzylidene-1-thio- $\beta$ -D-glucopyranoside (1) and ethyl 2,3,4-tri-O-benzyl-1-thio- $\alpha$ -D-mannopyranoside (5) in 5 steps. Building block 20 was obtained by coupling N-Z-protected 5-aminopentyl 2-O-benzyl-4,6-O-benzylidene- $\beta$ -D-glucopyranoside (14) obtained from the non-benzylated counterpart 12 with ethyl 2,3-di-O-benzoyl-4-O-chloroacetyl-1-thio- $\alpha$ -L-rhamnopyrano- side (18) obtained in 3 steps from ethyl 2,3-O-isopropylidene-1-thio- $\alpha$ -L-rhamnopyranoside (15).

In 1996, the first reports appeared that soil bacteria of the genus Arthrobacter can cause hitherto unknown severe infections in men. In this initial case, a patient who went through eye surgery developed a severe endophthalmitis caused by Arthrobacter.[1][2] Several other cases of Arthrobacter-associated infections have previously been reported including chronic uveitis, different forms of kryptogenic polyarthritis, seronegative spondilarthropathy, meningitis and lymphadenopathy (Whipple's syndrome). [3] Thus, synthetic fragments related to the exopolysaccharides of Arthobacter species are likely to be interesting targets because such oligosaccharide fragments could be used for diagnostic purposes or as artificial vaccines. Furthermore, exopolysaccharides of Arthrobacter are of significant importance as thickeners for the food industry or as stabilizers for foams and emulsions and as a flocculant.

Of the various *Arthrobacter* exopolysaccharides<sup>[4–9]</sup> studied so far only a few structures have been determined in detail.<sup>[7–9]</sup> As part of a project toward the synthesis of pyruvated oligosaccharides<sup>[10]</sup> we were especially interested in Simusan,<sup>[9]</sup> an acidic exopolysaccharide of *Arthrobacter sp.*, strain CE-17 which exhibits an octasaccharide repeating unit that contains a 4,6-pyruvated  $\beta$ -D-mannopyranosyl residue (Figure 1). In particular, the synthesis of a 5-aminopentyl glycoside tetrasaccharide related to the sequence **ABCD** of Simusan (Figure 1) that contains a  $\beta$ -D-mannopyranosyl residue is described here.

Figure 1. Repeating unit of the exopolysaccharide of *Arthrobacter sp.* CE-17

For the synthesis of the tetrasaccharide related to the sequence **ABCD** of Simusan we chose a blockwise approach. Thus, two disaccharide building blocks – β-D-Manp-(1 $\rightarrow$ 4)-D-Glcp as the disaccharide donor and  $\alpha$ -L-Rhap- $(1\rightarrow 3)$ -D-Glcp as the disaccharide acceptor – were needed. The donor block contains a β-D-mannosyl residue which was planned to be introduced by intramolecular glycosylation of a suitable prearranged glycoside. Recently, we developed a highly selective β-mannosylation procedure<sup>[11]</sup> which was applied to the preparation of the latter as follows. Starting from known [12] phenyl 4,6-O-benzylidene-1thio-β-D-glucopyranose (1), phase-transfer catalyzed benzoylation<sup>[13]</sup> afforded dibenzoate 2 (11%), 3-O-benzoate 3 (16%), and the desired 2-O-benzoate 4 (54%). Alternative selective benzoylations of 1 gave lower yields of compound 4 (see Experimental Section for details). Next, readily available<sup>[14]</sup> ethyl 2,3,4-tri-O-benzyl-1-thio-α-D-mannopyranoside (5) was esterified with *tert*-butyl malonate [15] (6) to give mannoside 7 (64%), the tert-butyl group of which was finally cleaved with trifluoroacetic acid to afford mannosyl malonate 8 in quantitative yield. The malonic acid tether in compound 8 was chosen for linking the mannose unit with glucoside 4 because previously, similarly tethered glycosides

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Scheme 1

gave high \(\beta\)-selectivities for intramolecular mannosylations.[11] Furthermore, combining the phenylthio group in the glucose moiety 4 with the ethylthio group in 8 allows us to make use of the principle of armed-disarmed glycosides<sup>[16]</sup> since mannosylation (i.e. activation of the ethylthio group) is possible without affecting the phenylthio group. Thus, according to previous procedures, [17][18] compounds 4 and 8 were condensed to give first saccharide 9 (81%), the benzylidene acetal of the glucose moiety of which was subsequently opened with NaCNBH3[17-19] to afford the prearranged glycoside 10 in 71% yield. As was expected from previous experiments,[11] intramolecular mannosylation of the latter with methyl trifluoromethanesulfonate (MeOTf) as the promotor proceeded smoothly to give exclusively the desired  $\beta$ -D-Manp-(1 $\rightarrow$ 4)-D-Glcp disaccharide donor block 11 in 69% yield. The β-linkage of the mannosyl moiety in 11 was evident from the NMR spectra of that compound which showed a typical<sup>[20-22]</sup> C,H-coupling constant at the anomeric center of 153.7 Hz.

The required  $\alpha$ -L-Rhap-(1 $\rightarrow$ 3)-D-Glcp disaccharide acceptor was prepared conventionally from 5-benzyloxy-carbonylpentyl 4,6-O-benzylidene- $\beta$ -D-glucopyranoside [23] (12). First, regioselective benzylation of the latter under phase-transfer conditions [24] afforded 13 (21%) and 14 (53%). Next, a properly protected rhamnose derivative was prepared from ethyl 2,3-O-isopropylidene-1-thio- $\alpha$ -L-rhamnopyranoside [25] (15) by chloroacetylation at the 4-position to give rhamnoside 16 (81%) followed by acidic hydrolysis of the isopropylidene group via 17 (88%) and rebenzoylation of the free OH-groups to give monosaccharide 18 (83%). Coupling of the latter to the 3-position of compound 14 was best performed with N-iodosuccinimide (NIS) in the presence of 0.5 equiv. of TMSOTf in dichloromethane.

Thus, the disaccharide 19 was obtained in 79% yield and the  $\alpha$ -linkage of the rhamnosyl group was once again proven by measuring the C,H coupling constant at the anomeric center of 174.4 Hz. Finally, the chloroacetyl group of compound 19 was removed with thiourea to afford the disaccharide acceptor 20 (87%).

Scheme 2

The coupling of disaccharide blocks 11 and 20 was somehow crucial. Several methods for the activation of thioglycosides including NIS/trifluoromethanesulfonic acid (TfOH), [26] bromine/silver trifluoromethanesulfonate (AgOTf), [27] and dimethyl (methylthio)sulfonium triflate (DMTST) [28] failed completely and resulted in products of decomposition. Only NIS/trimethylsilyl trifluoromethanesulfonate (TMSOTf) [29] gave the desired tetrasaccharide 21. However, the reaction showed an unusual temperature de-

Prearranged Glycosides, 9 FULL PAPER

pendence. At  $0^{\circ}$ C, compound 21 was obtained in poor 9% yield, whereas at  $-30^{\circ}$ C or  $-70^{\circ}$ C the yield of tetrasaccharide 21 could be enhanced to 32% and 64%, respectively. Final deblocking of the latter afforded the tetrasaccharide 5-aminopentyl glycoside (22) in 87% yield.

In summary, the efficient construction of the  $\beta$ -mannosyl-containing disaccharide block 11 using intramolecular glycosylation techniques in combination with the principle of armed and disarmed glycosyl donors demonstrated the usefulness of this appraoch for the construction of higher oligosaccharides. Furthermore, compound 11 can be used for the synthesis of other oligosaccharides containing this disaccharide fragment.

#### **Experimental Section**

General: Thin-layer chromatography (TLC) was performed on precoated plastic sheets, Polygram SIL G/UV<sub>254</sub>, 40 × 80 mm (Macherey-Nagel) using appropriately adjusted mixtures of toluene/acetone for the developing. Spots were detected by UV light and by charring with 5% sulfuric acid in ethanol. - Column chromatography (CC) was performed by elution from columns of silica gel S (Macherey-Nagel, 0.032-0.063 mm). - Solutions in organic solvents were dried with anhydrous sodium sulfate, and concentrated at 40°C, <200 Pa. - NMR spectra were recorded for solutions in CDCl<sub>3</sub> (with TMS as an internal standard) at 25°C on a Bruker AC 250 F spectrometer. Proton signal assignments were made by first order analysis of the spectra and by H,H-cosy techniques. Of two magnetically nonequivalent geminal protons, the one resonating at lower field was designated as Ha and the one resonating at higher field was designated as H<sub>b</sub>. Carbon signal assignments were made by C,H-correlation and by comparison of the peaks with those of related compounds. – Optical rotations were measured at 20°C with a Perkin-Elmer automatic polarimeter, Model 241. – Melting points were determined with a Büchi apparatus, Model SMP-20.

Phenyl 2,3-di-O-Benzoyl-4,6-O-benzylidene-1-thio-β-D-glucopyranoside (2), Phenyl 3-O-Benzoyl-4,6-O-benzylidene-1-thio-β-D-glucopyranoside (3), and Phenyl 2-O-Benzoyl-4,6-O-benzylidene-1-thio-β-Dglucopyranoside (4): A) Benzoyl chloride (165 µL, 1.39 mmol) was added at -50°C to a solution of 1<sup>[12]</sup> (0.50 g, 1.39 mmol) in pyridine (15 mL), the mixture was stirred for 48 h, poured into ice-cold water and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic phases were subsequently washed with aqueous HCl and NaHCO3 solution, dried and concentrated. Chromatography (gradient elution with toluene/acetone 55:1→35:1 v/v) afforded first 2 (116 mg, 15%), m.p. 203°C [ref. [30] 203–205°C (EtOH/ethyl acetate)].  $- [\alpha]_D^{20} =$ +42.6 (c = 1.1, CHCl<sub>3</sub>) {ref.<sup>[30]</sup> [ $\alpha$ ]<sub>D</sub> = +40 (c = 1.0, CHCl<sub>3</sub>)}. Eluted next was 3 (295 mg, 46%), m.p. 154-156°C (EtOH).  $[\alpha]_D = -98.4$  (c = 1.3, CHCl<sub>3</sub>).  $- {}^{1}H$  NMR (CDCl<sub>3</sub>):  $\delta = 5.51$  (s, 1 H, PhCH), 5.50 (dd,  $J_{3,4} = 9.5$  Hz, 1 H, 3-H), 4.76 (d,  $J_{1,2} =$ 9.7 Hz, 1 H, 1-H), 4.39 (dd,  $J_{5,6a} = 4.9$  Hz,  $J_{6a,6b} = -10.4$  Hz, 1 H, 6a-H), 3.81 (t,  $J_{5,6b} = 9.6$  Hz, 1 H, 6b-H), 3.77 (t,  $J_{4,5} = 9.6$  Hz, 1 H, 4-H), 3.70 (bd,  $J_{2,3} = 8.9$  Hz, 1 H, 2-H), 3.62 (dt, 1 H, 5-H), 3.04 (br. s, 1 H, OH). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 166.6$  (PhCO), 101.4 (PhCH), 89.2 (C-1), 78.2 (C-4), 75.6 (C-3), 70.7 (C-5), 68.5 (2 C, C-2, C-6). - C<sub>26</sub>H<sub>24</sub>O<sub>6</sub>S (464.5): calcd. C 67.23, H 5.21, S 6.90; found C 67.19, H 5.26, S 6.95.

Eluted next was unchanged 1 (69 mg, 14%).

**B)** A mixture of **1** (0.50 g, 1.39 mmol) and di-*n*-butyltin oxide (0.36 g, 1.45 mmol) in benzene (100 mL) was heated for 24 h under

reflux in a Dean-Stark apparatus, cooled to room temperature and concentrated. The residue was dissolved in benzene (50 mL), a solution of benzoyl chloride (0.18 mL, 1.53 mmol) in benzene (25 mL) was added dropwise at room temperature and the solution was stirred for 24 h. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, filtered through a layer of Celite® and concentrated. Chromatography as described above afforded first 3 (0.38 g, 59%). Eluted next was 4 (0.15 g, 23%), which was obtained as a colorless foam.  $- [\alpha]_D^{20} =$ -27.1 (c = 0.7, CHCl<sub>3</sub>). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 5.54 (s, 1 H, PhCH), 5.15 (dd,  $J_{2,3} = 8.8$  Hz, 1 H, 2-H), 4.88 (d,  $J_{1,2} = 10.0$  Hz, 1 H, 1-H), 4.40 (dd,  $J_{5,6a} = 4.6$  Hz,  $J_{6a,6b} = -10.5$  Hz, 1 H, 6a-H), 4.03 (t,  $J_{3,4} = 8.9$  Hz, 1 H 3-H), 3.81 (t,  $J_{5,6b} = 10.0$  Hz, 1 H, 6b-H), 3.64-3.58 (m,  $J_{4,5} = 9.3$  Hz, 2 H, 4-H, 5-H), 2.85 (br. s, 1 H, OH).  $- {}^{13}$ C NMR (CDCl<sub>3</sub>):  $\delta = 165.8$  (PhCO), 101.9 (PhCH), 86.6 (C-1), 80.6 (C-4), 73.6 (C-3), 73.2 (C-2), 70.4 (C-5), 68.5 (C-6). - C<sub>26</sub>H<sub>24</sub>O<sub>6</sub>S (464.5): calcd. C 67.23, H 5.21, S 6.90; found C 67.17, H 5.08, S 7.04.

C) An aqueous NaOH solution (5%, 32 mL) was added with vigurous stirring  $-5^{\circ}$ C to a solution of **1** (4.54 g, 12.6 mmol), Bu<sub>4</sub>NHSO<sub>4</sub> (0.83 g, 2.58 mmol) and benzoyl chloride (1.97 mL, 17.02 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (230 mL). The mixture was stirred for 10 min, the organic layer was separated, washed with water, dried and concentrated. Chromatography as described above afforded first **2** (0.77 g, 11%), then **3** (0.96 g, 16%), and finally **4** (3.14 g, 54%).

Ethyl 2,3,4-Tri-O-benzyl-6-O-(tert-butyloxycarbonylethanoyl)-1-thio- $\alpha$ -D-mannopyranoside (7): DCC (0.96 g, 4.65 mmol) was added at  $0^{\circ}$ C to a solution of  $5^{[14]}$  (2.09 g, 4.23 mmol),  $6^{[15]}$  (0.75 g, 4.65 mmol) and 1-hydroxy-1H-benzotriazole (0.69 g, 5.08 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL). The mixture was stirred at room temperature for 24 h, diluted with CH<sub>2</sub>Cl<sub>2</sub> and filtered through a layer of Celite®. The filtrate was subsequently washed with aqueous HCl and NaHCO<sub>3</sub> solution, dried and concentrated. Chromatography (nhexane/ethyl acetate 6:1 v/v) of the residue afforded 7 (1.73 g, 64%) as a colorless foam.  $- [\alpha]_D^{20} = +58.0 \ (c = 1.1, CHCl_3). - {}^{1}H$ NMR (CDCl<sub>3</sub>):  $\delta = 5.33$  (d,  $J_{1,2} = 1.0$  Hz, 1 H, 1-H), 4.92 (d, J =-10.7 Hz, 1 H, PhCH<sub>2</sub>), 4.71 (d, J = -12.5 Hz, 1 H, PhCH<sub>2</sub>), 4.65  $(d, J = -12.5 \text{ Hz}, 1 \text{ H}, PhCH_2), 4.60-4.56 \text{ (m}, 3 \text{ H}, PhCH_2), 4.44$  $(dd, J_{5,6a} = 5.1 \text{ Hz}, J_{6a,6b} = -11.8 \text{ Hz}, 1 \text{ H}, 6a-\text{H}), 4.36 (dd, J_{5,6b} = -11.8 \text{ Hz})$ 2.2 Hz, 1 H, 6b-H), 4.18 (ddd, 1 H, 5-H), 3.93 (t,  $J_{4.5} = 9.4$  Hz, 1 H, 4-H), 3.83 (2 dd,  $J_{2,3} = 3.2$  Hz,  $J_{3,4} = 10.1$  Hz, 2 H, 2-H, 3-H), 3.29 (s, 2 H, COCH<sub>2</sub>), 2.66-2.47 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.44 [s, 9 H,  $C(CH_3)_3$ ], 1.23 (t, J = 7.4 Hz, 3 H,  $SCH_2CH_3$ ).  $- {}^{13}C \text{ NMR}$ (CDCl<sub>3</sub>):  $\delta = 166.9$  (CO), 165.4 (CO), 82.0 (C(CH<sub>3</sub>)<sub>3</sub>), 81.9 (C-1), 80.2 (C-3), 76.2 (C-2), 75.2 (PhCH<sub>2</sub>), 74.6 (C-4), 72.0 (2 C, PhCH<sub>2</sub>), 70.1 (C-5), 64.3 (C-6), 42.6 (COCH<sub>2</sub>), 27.9 (C(CH<sub>3</sub>)<sub>3</sub>), 25.4  $(SCH_2CH_3)$ , 14.9  $(SCH_2CH_3)$ . -  $C_{36}H_{44}O_8S$  (636.8): calcd. C 67.90, H 6.96, S 5.04; found C 67.94, H 7.08, S 5.11.

Ethyl 2,3,4-Tri-O-benzyl-6-O-(hydroxycarbonylethanoyl)-1-thio- $\alpha$ -D-mannopyranoside (8): Trifluoroacetic acid (5.3 mL, 51.8 mmol) was added at room temperature to a solution of 7 (1.65 g, 2.59 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (80 mL), the mixture was stirred for 3 h and concentrated. Coevaporation of toluene (3 times) afforded crude 8 (2.58 g, 100%) which was used without further purification in the next step.

Ethyl 2,3,4-tri-O-benzyl-6-O-[(2-O-benzyl-4,6-O-benzylidene-1-phenylthio- $\beta$ -D-glucopyranos-3-yloxy)carbonylethanoyl]-1-thio- $\alpha$ -D-mannopyranoside (9): DCC (0.47 g, 2.28 mmol) was added at room temperature to a solution of 4 (1.04 g, 2.24 mmol), crude 8 (1.19 g, 2.05 mmol) and a catalytic amount of DMAP (ca. 20 mg) in CH<sub>2</sub>Cl<sub>2</sub> (30 mL), and the mixture was stirred for 24 h. Workup as described for the preparation of 7 and chromatography (toluene/ethyl acetate 22:1 v/v) afforded 9 (1.86 g, 81%) as a colorless foam.

FULL PAPER \_\_\_\_\_ G. Lemanski, T. Ziegler

 $- [\alpha]_D^{20} = +36.8 \ (c = 1.1, CHCl_3). - {}^{1}H \ NMR \ (CDCl_3): \delta =$ 5.54 (t,  $J_{3,4} = 9.3$  Hz, 1 H, 3-H), 5.49 (s, 1 H, PhCH), 5.28 (dd,  $J_{2,3} = 9.1 \text{ Hz}, 1 \text{ H}, 2\text{-H}, 5.27 \text{ (s, 1 H, 1'-H)}, 4.93 \text{ (d, } J_{1,2} = 10.0 \text{ Hz},$ 1 H, 1-H), 4.85 (d, J = -10.9 Hz, 1 H, PhCH<sub>2</sub>), 4.68 (d, J =-12.1 Hz, 1 H, PhCH<sub>2</sub>), 4.62 (d,, J = -12.3 Hz 1 H, PhCH<sub>2</sub>), 4.54 (s, 2 H, PhCH<sub>2</sub>), 4.49 (d, J = -10.9 Hz, 1 H, PhCH<sub>2</sub>), 4.41 (dd,  $J_{5,6a} = 4.7 \text{ Hz}, J_{6a,6b} = -10.4 \text{ Hz}, 1 \text{ H}, 6a-H), 4.25-4.20 \text{ (m, 2 H, 1)}$ 6a'-H, 6b'-H), 4.10-4.04 (m, 1 H, 5'-H), 3.83-3.76 (m,  $J_{5,6b}$  = 9.6 Hz, 4 H, 2'-H, 3'-H, 4'-H, 6b-H), 3.75 (t,  $J_{4.5} = 9.6$  Hz, 1 H, 4-H), 3.63 (dt, 1 H, 5-H), 3.27 (s, 2 H, COCH<sub>2</sub>), 2.60-2.43 (m, 2 H,  $SCH_2CH_3$ ), 1.19 (t, J = 7.4 Hz, 3 H,  $SCH_2CH_3$ ).  $- {}^{13}C \text{ NMR}$ (CDCl<sub>3</sub>):  $\delta = 165.4$  (CO), 165.3 (CO), 165.2 (CO), 101.4 (PhCH), 87.0 (C-1), 81.8 (C-1'), 80.2 (C-3'), 78.0 (C-4), 76.1, 74.6 (C-2',4'), 75.0 (PhCH<sub>2</sub>), 73.6 (C-3), 72.0 (2 C, PhCH<sub>2</sub>), 70.8 (2 C, C-2, C-5), 70.0 (C-5'), 68.4 (C-6), 64.4 (C-6'), 40.8 (COCH<sub>2</sub>), 25.3  $(SCH_2CH_3)$ , 14.9  $(SCH_2CH_3)$ . -  $C_{58}H_{58}O_{13}S_2$  (1027.2): calcd. C 67.82, H 5.69, S 6.24; found C 67.77, H 5.73, S 6.22.

2,3,4-tri-O-benzyl-6-O-[(2-O-benzoyl-6-O-benzyl-1-phenylthio-β-D-glucopyranos-3-yloxy)carbonylethanoyl]-1-thio-α-D-mannopyranoside (10): A solution of HCl in THF was added portionwise at room temperature to a suspension of 9 (1.60 g, 1.51 mmol), NaCNBH<sub>3</sub> (1.19 g, 18.9 mmol) and 3 Å molecular sieves (ca. 1.0 g) in THF (50 mL) until the evolution of gas ceased. The mixture was diluted with CH2Cl2 and filtered through a layer of Celite®. The filtrate was washed with an aqueous NaHCO3 solution, dried and concentrated. Chromatography (toluene/ethyl acetate 8:1 v/v) afforded **10** (1.10 g, 71%) as a colorless foam.  $- [\alpha]_D^{20} = +42.5$  (c = 1.6, CHCl<sub>3</sub>). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 5.30$  (s, 1 H, 1'-H), 5.30-5.25 (m,  $J_{3,4} = 8.9$  Hz, 1 H, 3-H), 5.13 (t,  $J_{2,3} = 9.7$  Hz, 1 H, 2-H), 4.89 (d, J = -11.0 Hz, 1 H, PhCH<sub>2</sub>), 4.81 (d,  $J_{1,2} =$ 9.9 Hz, 1 H, 1-H), 4.68 (d, J = -12.3 Hz, 1 H, PhCH<sub>2</sub>), 4.55 (br. s, 5 H, PhCH<sub>2</sub>), 4.53 (d, J = -11.3 Hz, 1 H, PhCH<sub>2</sub>), 4.37-4.27 (m, 2 H, 6a'-H, 6b'-H), 4.15-4.10 (m, 1 H, 5'-H), 3.92 (t,  $J_{4'.5'}$  = 9.2 Hz, 1 H, 4'-H), 3.84 (m, 3 H, 2'-H, 3'-H, 6a-H), 3.73-3.66 (m, 2 H, 6b-H, OH), 3.61 (bm, 2 H, 4-H, 5-H), 3.35 (d, J = -16.1 Hz, 1 H, COCH<sub>2</sub>), 3.20 (d, J = -16.1 Hz, 1 H, COCH<sub>2</sub>), 2.59-2.46 (m, 2 H,  $SCH_2CH_3$ ), 1.20 (t, J = 7.4 Hz, 3 H,  $SCH_2CH_3$ ).  $- ^{13}C$ NMR (CDCl<sub>3</sub>):  $\delta = 166.9$  (PhCO), 165.6 (COCH<sub>2</sub>), 165.2 (COCH<sub>2</sub>), 85.9 (C-1), 81.9 (C-1'), 80.0 (C-3'), 79.4 (C-5), 78.2 (C-3), 76.2 (C-2'), 75.1 (PhCH<sub>2</sub>), 74.0 (C-4'), 73.5 (PhCH<sub>2</sub>), 72.0 (PhCH<sub>2</sub>), 71.9 (PhCH<sub>2</sub>), 70.2 (C-2), 69.8 (C-5'), 69.4 (C-6), 68.9 (C-4), 64.6 (C-6'), 41.3 (COCH<sub>2</sub>), 25.5 (SCH<sub>2</sub>CH<sub>3</sub>), 14.9  $(SCH_2CH_3)$ . -  $C_{58}H_{60}O_{13}S_2$  (1029.2): calcd. C 67.69, H 5.88, S 6.23; found C 67.57, H 5.88, S 6.21.

O-(2,3,4-Tri-O-benzyl-β-D-mannopyranosyl)-(1 $\rightarrow$ 4)-2-Obenzoyl-6-*O*-benzyl-1-thio-β-D-glucopyranoside 3,6'-Malonate (11): MeOTf (0.37 mL, 3.40 mmol) was added at room temperature under argon to a mixture of 10 (0.70 g, 0.68 mmol) and 4 Å molecular sieves (1.76 g) in acetonitrile (30 mL). The mixture was stirred for 6 h, diluted with CH<sub>2</sub>Cl<sub>2</sub> and filtered. The filtrate was washed with water, dried and concentrated. Chromatography (toluene/acetone 40:1 v/v) of the residue afforded 11 (0.45 g, 69%) as a colorless foam.  $- [\alpha]_D^{20} = -4.2$  (c = 1.0, CHCl<sub>3</sub>).  $- {}^{1}$ H NMR (CDCl<sub>3</sub>):  $\delta = 5.31$  (t,  $J_{3.4} = 9.1$  Hz, 1 H, 3-H), 5.15 (t,  $J_{2.3} = 9.6$  Hz, 1 H, 2-H), 4.86 (d, J = -11.3 Hz, 1 H, PhCH<sub>2</sub>), 4.82 (d,  $J_{1,2} = 10.1$  Hz, 1 H, 1-H), 4.74 (d, J = -11.8 Hz, 1 H, PhCH<sub>2</sub>), 4.70 (d, J =-12.5 Hz, 1 H, PhCH<sub>2</sub>),  $4.56-4.50 \text{ (m, } J_{6a',6b'} = -11.1 \text{ Hz, 1 H,}$ 6a'-H), 4.50 (s, 3 H, PhCH<sub>2</sub>), 4.47 (d, J = -11.8 Hz, 1 H, PhCH<sub>2</sub>),  $4.36 \text{ (d, } J = -12.1 \text{ Hz, } 1 \text{ H, PhCH}_2), 4.29 \text{ (s, } 1 \text{ H, } 1'\text{-H)}, 4.03 \text{ (dd, } 1 \text{ Hz, } 1'\text{-Hz})$  $J_{5',6b'} = 8.4 \text{ Hz}, 1 \text{ H}, 6b'-\text{H}), 3.97 \text{ (t}, J_{4,5} = 9.3 \text{ Hz}, 1 \text{ H}, 4-\text{H}), 3.74$ (t,  $J_{4',5'}$  = 9.2 Hz, 1 H, 4'-H), 3.60 (bd,  $J_{2',3'}$  = 2.9 Hz,  $J_{5,6a}$  = 10.0 Hz,  $J_{6a,6b} = -11.0 \text{ Hz}$ , 3 H, 2'-H, 5'-H, 6a-H), 3.53 (bt,  $J_{5,6b} = 3.7 \text{ Hz}, 1 \text{ H}, 5\text{-H}), 3.44 \text{ (dd}, 1 \text{ H}, 6b\text{-H}), 3.38 \text{ (dd}, <math>J_{3',4'} =$ 

8.8 Hz, 3'-H), 3.17 (bd, 2 H, COCH<sub>2</sub>). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  = 165.6, 165.2, 164.5 (CO), 103.5 (C-1',  $J_{\text{C-1',1'-H}}$  = 153.7 Hz), 85.5 (C-1,  $J_{\text{C-1,1-H}}$  = 157.0 Hz), 82.2 (C-3'), 78.9 (C-5), 78.6 (C-4), 76.7 (2 C, C-3, C-4'), 74.8 (PhCH<sub>2</sub>), 74.4 (C-2'), 73.7, 73.5, 72.0 (PhCH<sub>2</sub>), 71.2 (C-2), 70.4 (C-5'), 68.4 (C-6), 64.2 (C-6'), 42.7 (COCH<sub>2</sub>). - C<sub>56</sub>H<sub>54</sub>O<sub>13</sub>S (967.1): calcd. C 69.55, H 5.63, S 3.32; found C 69.26, H 5.41, S 3.26.

5-Benzyloxycarbonylaminopentyl 3-O-Benzyl-4,6-O-benzylidene-β-D-glucopyranoside (13) and 5-Benzyloxycarbonylaminopentyl 2-O-Benzyl-4,6-*O*-benzylidene-β-D-glucopyranoside (14): Aqueous 5% NaOH solution (7.95 mL) was added with vigurous stirring at room temperature to a solution of 12<sup>[23]</sup> (2.79 g, 5.72 mmol), BnBr (1.19 mL, 10.02 mmol) and Bu<sub>4</sub>NHSO<sub>4</sub> (0.38 g, 1.13 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and the mixture was refluxed for 72 h. The mixture was diluted with CH2Cl2, washed with water, dried and concentrated. Chromatography (gradient elution with toluene/acetone  $15:1\rightarrow 10:1$  v/v) of the residue afforded first 14 (1.82 g, 53%), m.p. 112–113 °C (EtOH). –  $[\alpha]_D^{20} = -19.3$  (c = 1.6, CHCl<sub>3</sub>). –  ${}^{1}$ H NMR (CDCl<sub>3</sub>):  $\delta = 5.49$  (s, 1 H, PhCH), 5.07 (s, 2 H,  $PhCH_2OCO$ ), 4.92 (d, J = -11.5 Hz, 1 H,  $PhCH_2$ ), 4.73 (bd, J = -11.5 Hz) -11.5 Hz, 2 H, PhCH<sub>2</sub>, NH), 4.48 (d,  $J_{1,2} = 7.7 \text{ Hz}$ , 1 H, 1-H), 4.32 (dd,  $J_{5,6a} = 4.9$  Hz,  $J_{6a,6b} = -10.4$  Hz, 1 H, 6a-H), 3.93-3.87 (m, 1 H, OCH<sub>2</sub>), 3.85-3.79 (m,  $J_{3,4} = 9.1$  Hz,  $J_{3,OH} = 2.1$  Hz, 1 H, 3-H), 3.75 (t,  $J_{5,6b} = 10.2$  Hz, 1 H, 6b-H), 3.58-3.53 (m, 1 H, OCH<sub>2</sub>), 3.52 (t,  $J_{4,5} = 9.3$  Hz, 1 H, 4-H), 3.43-3.37 (m,  $J_{5,6a} =$ 4.9 Hz, 1 H, 5-H), 3.33 (dd,  $J_{2,3} = 8.8$  Hz, 1 H, 2-H), 3.17-3.11 (m, 2 H, CH<sub>2</sub>), 2.59 (d, 1 H, OH), 1.66-1.33 (m, 6 H, CH<sub>2</sub>). -<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 156.3$  (OCONH), 103.7 (C-1), 101.7 (PhCH), 81.8 (C-2), 80.4 (C-4), 74.7 (PhCH<sub>2</sub>), 73.1 (C-3), 70.1 (OCH<sub>2</sub>), 68.7 (C-6), 66.5 (PhCH<sub>2</sub>OCO), 66.0 (C-5), 40.8 (CH<sub>2</sub>NH), 29.6, 29.3, 23.2 (CH<sub>2</sub>). - C<sub>33</sub>H<sub>39</sub>NO<sub>8</sub> (577.7): calcd. C 68.61, H 6.81, N 2.43; found C 68.58, H 6.79, N 2.37.

Eluted next was **13** (0.68 g, 21%), m.p. 119–120°C (EtOH). –  $[\alpha]_D^{20} = -26.0$  (c = 1.8, CHCl<sub>3</sub>). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 5.54$  (s, 1 H, PhCH), 5.09 (s, 2 H, PhCH<sub>2</sub>OCO), 4.95 (d, J = -11.7 Hz, 1 H, PhCH<sub>2</sub>), 4.85 (br. s, 1 H, NH), 4.80 (d, J = -11.8 Hz, 1 H, PhCH<sub>2</sub>), 4.37 (d,  $J_{1,2} = 7.7$  Hz, 1 H, 1-H), 4.33 (dd,  $J_{5,6a} = 5.0$  Hz,  $J_{6a,6b} = -10.4$  Hz, 1 H, 6a-H), 3.92–3.84 (m, 1 H, OCH<sub>2</sub>), 3.78 (t,  $J_{5,6b} = 10.3$  Hz, 1 H, 6b-H), 3.72–3.66 (m, 1 H, 4-H), 3.65 (t,  $J_{3,4} = 8.7$  Hz, 1 H, 3-H), 3.58–3.48 (m, 2 H, 2-H, OCH<sub>2</sub>), 3.46–3.88 (m, 1 H, 5-H), 3.19–3.15 (m, 2 H, CH<sub>2</sub>), 2.70 (br. s, 1 H, OH), 1.69–1.36 (m, 6 H, CH<sub>2</sub>). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 156.4$  (OCONH), 103.3 (C-1), 101.2 (PhCH), 81.3 (C-3), 80.3 (C-4), 74.5 (PhCH<sub>2</sub>), 74.3 (C-2), 68.7 (C-6), 66.6 (PhCH<sub>2</sub>OCO), 66.4 (C-5), 40.8 (CH<sub>2</sub>NH), 29.5, 20.9, 23.0 (CH<sub>2</sub>). - C<sub>33</sub>H<sub>39</sub>NO<sub>8</sub> (577.7): calcd. C 68.61, H 6.81, N 2.43; found C 68.52, H 6.84, N 2.36.

Ethyl 4-*O*-Chloroacetyl-2,3-*O*-isopropylidene-1-thio-α-L-rhamnopyranoside (16): Chloroacetyl chloride (4.7 mL, 59.3 mmol) was added dropwise at 0°C to a solution of  $15^{[25]}$  (9.2 g, 37.1 mmol) and pyridine (4.0 mL, 49.6 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (180 mL). The mixture was stirred for 16 h at room temperature and poured into water. The organic layer was separated, washed with aqueous HCl and NaHCO<sub>3</sub> solution, dried and concentrated. Crystallisation of the residue from acetone afforded 16 (9.74 g, 81%), m.p. 122°C. – [α]<sub>D</sub><sup>20</sup> = -43.1 (c = 1.0, CHCl<sub>3</sub>). – <sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 5.56 (s, 1 H, 1-H), 4.96 (dd,  $J_{4,5}$  = 10.1 Hz, 1 H, 4-H), 4.22–4.19 (m, 1 H, 2-H), 4.18–4.12 (m,  $J_{3,4}$  = 7.5 Hz,  $J_{5,6}$  = 6.3 Hz, 2 H, 3-H, 5-H), 4.11 (2 s, 2 H, CICH<sub>2</sub>CO), 2.74–2.49 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.57 [s, 3 H, C(CH<sub>3</sub>)<sub>2</sub>], 1.34 [s, 3 H, C(CH<sub>3</sub>)<sub>2</sub>], 1.31 (t, J = 7.5 Hz, 3 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.19 (d, 1 H, 6-H). – <sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 166.6 (CICH<sub>2</sub>CO), 110.0 [C(CH<sub>3</sub>)<sub>2</sub>], 79.4 (C-1), 76.9, 76.8 (C-2,4), 75.2

Prearranged Glycosides, 9 FULL PAPER

(C-3), 64.2 (C-5), 27.7 [C( $CH_3$ )<sub>2</sub>], 26.4 [C( $CH_3$ )<sub>2</sub>], 24.5 (S $CH_2CH_3$ ), 16.9 (C-6), 14.6 (SCH<sub>2</sub> $CH_3$ ). – C<sub>13</sub>H<sub>21</sub>ClO<sub>5</sub>S (324.8): calcd C 48.07, H 6.52, Cl 10.92, S 9.87; found C 48.05, H 6.56, Cl 10.94, S 9.79

Ethyl 4-O-Chloroacetyl-1-thio-α-L-rhamnopyranoside (17): Acetic acid (150 mL) was added to a solution of 16 (8.39 g, 25.82 mmol) in a small amount of CH<sub>2</sub>Cl<sub>2</sub>. The mixture was stirred for 7 h at 60°C, cooled to room temperature and concentrated. Crystallisation of the residue from n-hexane/acetone afforded 17 (6.47 g. 88%), m.p. 137-139°C.  $- [\alpha]_D^{20} = -89.5$  (c = 0.7, CHCl<sub>3</sub>).  $- {}^{1}$ H NMR ([D<sub>6</sub>]acetone):  $\delta = 5.25$  (d,  $J_{1,2} = 1.0$  Hz, 1 H, 1-H), 4.99 (t,  $J_{4,5} = 9.7 \text{ Hz}, 1 \text{ H}, 4\text{-H}, 4.49 (d, J_{2,OH} = 4.1 \text{ Hz}, 1 \text{ H}, OH), 4.34$ (d, J = -15.0 Hz, 1 H, ClCH<sub>2</sub>CO), 4.26 (d, J = -15.0 Hz, 1 H,  $CICH_2CO$ ), 4.16 (d,  $J_{3,OH} = 7.0 \text{ Hz}$ , 1 H, OH), 4.13-4.03 (m,  $J_{5,6} = 6.3 \text{ Hz}, 1 \text{ H}, 5 \text{-H}), 3.97 \text{ (dt, } J_{2,3} = 3.4 \text{ Hz}, 1 \text{ H}, 2 \text{-H}), 3.78$ (ddd,  $J_{3,4} = 9.6 \text{ Hz}$ , 1 H, 3-H), 2.70-2.52 (m, 2 H, SC $H_2$ CH<sub>3</sub>), 1.25 (t, J = 7.4 Hz, 3 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.14 (d, 1 H, 6-H).  $- {}^{13}\text{C}$ NMR ([D<sub>6</sub>]acetone):  $\delta = 167.8$  (ClCH<sub>2</sub>CO), 85.4 (C-1), 77.3 (C-4), 73.4 (C-2), 70.5 (C-3), 67.0 (C-5), 41.7 (SCH<sub>2</sub>CH<sub>3</sub>), 17.6 (C-6), 15.3 (SCH<sub>2</sub>CH<sub>3</sub>). - C<sub>10</sub>H<sub>17</sub>ClO<sub>5</sub>S (284.8): calcd C 42.18, H 6.02, Cl 12.45, S 11.26; found C 42.33, H 5.99, Cl 12.97, S 11.16.

Ethyl 2,3-Di-O-benzoyl-4-O-chloroacetyl-1-thio-α-L-rhamnopyranoside (18): A mixture of BzCl (11.7 mL, 100.6 mmol) and pyridine (8.0 mL, 98.59 mmol) was slowly dropped at 0°C into a solution of 17 (5.73 g, 20.12 mmol) in acetonitrile (80 mL). The mixture was stirred for 2 h at room temperature, poured into water and extracted with CH2Cl2. The combined extracts were washed with aqueous HCl and NaHCO3 solution, dried and concentrated. Chromatography (n-hexane/ethyl acetate 12:1 v/v) of the residue and crystallisation from *n*-hexane/diethyl ether afforded **18** (8.23 g, 83%), m.p. 73-75°C.  $- [\alpha]_D^{20} = +31.9$  (c = 1.5, CHCl<sub>3</sub>).  $- {}^{1}$ H NMR (CDCl<sub>3</sub>):  $\delta = 5.73$  (dd,  $J_{2,3} = 3.3$  Hz, 1 H, 2-H), 5.61 (dd,  $J_{3,4} = 10.1 \text{ Hz}, 1 \text{ H}, 3\text{-H}), 5.49 \text{ (t, } J_{4,5} = 9.8 \text{ Hz}, 1 \text{ H}, 4\text{-H}), 5.45$ (d,  $J_{1,2} = 1.4$  Hz, 1 H, 1-H), 4.48-4.41 (m,  $J_{5,6} = 6.2$  Hz, 1 H, 5-H), 4.01 (d, J = -14.5 Hz, 1 H, ClCH<sub>2</sub>CO), 3.94 (d, J = -14.5 Hz, 1 H, CICH<sub>2</sub>CO), 2.78-2.64 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.35 (t, J =7.4 Hz, 3 H,  $SCH_2CH_3$ ), 1.35 (d, 1 H, 6-H).  $- {}^{13}C$  NMR (CDCl<sub>3</sub>):  $\delta = 166.7 \text{ (CICH}_2\text{CO)}, 165.4 \text{ (PhCO)}, 165.3 \text{ (PhCO)}, 82.0 \text{ (C-1)},$ 73.4 (C-4), 72.5 (C-2), 70.2 (C-3), 66.7 (C-5), 40.5 (ClCH<sub>2</sub>CO), 25.6  $(SCH_2CH_3)$ , 17.4 (C-6), 14.9  $(SCH_2CH_3)$ . –  $C_{24}H_{25}CIO_7S$  (493.0): calcd C 58.47, H 5.11, S 6.50; found C 58.49, H 5.03, S 6.56.

5-Benzyloxycarbonylaminopentyl O-(2,3-Di-O-benzoyl-4-O-chloroacetyl-α-L-rhamnopyranosyl)-(1→3)-2-O-benzyl-4,6-O-benzylidene-β-D-glucopyranoside (19): A mixture of 14 (0.98 g, 1.70 mmol), **18** (1.01 g, 2.04 mmol) and 4 Å molecular sieves (ca. 1.0 g) in CH<sub>2</sub>Cl<sub>2</sub> (25 mL) was cooled under argon to 0°C and stirred for 10 min. NIS (0.46 g, 2.04 mmol) and TMSOTf (184 µL, 1.02 mmol) were successively added, the mixture was stirred for 20 min, neutralized by the addition of triethylamine and filtered through a layer of Celite®. The filtrate was washed with aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution, dried and concentrated. Chromatography (toluene/acetone 17:1 v/v) of the residue afforded 19 (1.35 g, 79%) as a colorless foam.  $- [\alpha]_D^{20} = +38.4$  (c = 1.2, CHCl<sub>3</sub>).  $- {}^{1}H$  NMR (CDCl<sub>3</sub>):  $\delta = 5.71$  (dd,  $J_{2',3'} = 3.5$  Hz, 1 H, 2'-H), 5.64 (dd,  $J_{3',4'} =$ 10.0 Hz, 1 H, 3'-H), 5.55 (s, 1 H, PhCH), 5.35 (d,  $J_{1',2'} = 1.3$  Hz, 1 H, 1'-H), 5.27 (t,  $J_{4',5'} = 10.0$  Hz, 1 H, 4'-H), 5.08 (br. s, 2 H,  $PhCH_2OCO$ ), 4.89 (d, J = -10.7 Hz, 1 H,  $PhCH_2$ ), 4.73 (bd, 2 H, PhCH<sub>2</sub>, NH), 4.49 (d,  $J_{1,2} = 7.7$  Hz, 1 H, 1-H), 4.42-4.32 (m,  $J_{5',6'} = 6.2 \text{ Hz}, 2 \text{ H}, 5'-\text{H}, 6a-\text{H}), 3.96 \text{ (t, } J_{3,4} = 9.2 \text{ Hz}, 1 \text{ H}, 3-\text{H}),$ 3.90-3.88 (m, 1 H, OCH<sub>2</sub>), 3.84-3.77 (m, 2 H, 6b-H), 3.83 (d,  $J = -14.5 \text{ Hz}, 1 \text{ H}, \text{ ClCH}_2\text{CO}), 3.78 \text{ (d, } J = -14.5 \text{ Hz}, 1 \text{ H},$ CICH<sub>2</sub>CO), 3.64 (t,  $J_{4,5} = 9.5$  Hz, 1 H, 4-H), 3.60-3.50 (m, 2 H,

OCH<sub>2</sub>, 2-H), 3.48-3.40 (m, 1 H, 5-H), 3.19-3.12 (m, 2 H, CH<sub>2</sub>), 1.68-1.64 (m, 2 H, CH<sub>2</sub>), 1.54-1.37 (m, 4 H, CH<sub>2</sub>), 0.86 (d, 1 H, 6'-H). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  = 166.6, 165.6, 165.2 (CO), 156.4 (OCONH), 104.2 (C-1), 101.9 (PhCH), 97.9 (C-1',  $J_{\text{C-1',1'-H}}$  = 174.4 Hz), 82.7 (C-2), 79.1 (C-4), 76.4 (C-3), 75.2 (PhCH<sub>2</sub>), 73.3 (C-4'), 70.4 (C-2'), 70.2 (OCH<sub>2</sub>), 70.0 (C-3'), 68.8 (C-6), 66.6 (PhCH<sub>2</sub>OCO), 66.4 (C-5), 65.8 (C-5'), 40.9 (CH<sub>2</sub>NH), 40.4 (ClCH<sub>2</sub>CO), 29.7 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 23.3 (CH<sub>2</sub>), 16.8 (C-6'). -C<sub>55</sub>H<sub>58</sub>ClNO<sub>15</sub> (1008.5): calcd C 65.50, H 5.80, N 1.39; found C 65.41, H 5.93, N 1.30.

5-Benzyloxycarbonylaminopentyl *O*-(2,3-Di-*O*-benzoyl-α-L-rhamnopyranosyl)-(1→3)-2-O-benzyl-4,6-O-benzylidene-β-D-glucopyranoside (20): A solution of 19 (0.60 g, 0.6 mmol) and thiourea (0.2 g, 2.63 mmol) in a mixture of MeOH (17 mL) and CH<sub>2</sub>Cl<sub>2</sub> (6 mL) was stirred at room temperature for 3 d and concentrated. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>, washed with water, dried and concentrated. Chromatography (toluene/acetone 12:1 v/v) of the residue afforded **20** (0.48 g, 87%) as a colorless foam.  $- [\alpha]_D^{20} =$ +17.4 (c = 1.0, CHCl<sub>3</sub>). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 5.67$  (dd,  $J_{2',3'} = 3.4 \text{ Hz } 1 \text{ H}, 2'-\text{H}), 5.56 \text{ (s, 1 H, PhCH)}, 5.50 \text{ (dd, } J_{3',4'} =$ 9.9 Hz, 1 H, 3'-H), 5.33 (d,  $J_{1',2'} = 1.4$  Hz, 1 H, 1'-H), 5.07 (s, 2 H, PhCH<sub>2</sub>OCO), 4.89 (d, J = -10.7 Hz, 1 H, PhCH<sub>2</sub>), 4.73 (bd, 2 H, PhCH<sub>2</sub>, NH), 4.49 (d,  $J_{1,2} = 7.7$  Hz, 1 H, 1-H), 4.36 (dd,  $J_{5,6a} =$ 4.9 Hz,  $J_{6a,6b} = -10.3$  Hz, 1 H, 6a-H), 4.21-4.16 (m,  $J_{5',6'} =$ 6.1 Hz, 1 H, 5'-H), 3.97 (t,  $J_{3,4} = 9.2$  Hz, 1 H, 3-H), 3.95-3.88 (m, 1 H, OCH<sub>2</sub>), 3.80 (t,  $J_{5,6b}$  = 9.6 Hz, 1 H, 6b-H), 3.75 (dt,  $J_{4'.5'}$  = 9.7 Hz,  $J_{4',OH} = 5.5$  Hz, 1 H, 4'-H), 3.63 (t,  $J_{4.5} = 9.4$  Hz, 1 H, 4-H), 3.57-3.50 (m, 2 H, OCH<sub>2</sub>, 2-H), 3.44 (dt, 1 H, 5-H), 3.16-3.11 (m, 2 H, CH<sub>2</sub>), 2.13 (d, 1 H, OH), 1.68-1.62 (m, 2 H, CH<sub>2</sub>), 1.53-1.36 (m, 4 H, CH<sub>2</sub>), 1.02 (d, 1 H, 6'-H). - <sup>13</sup>C NMR  $(CDCl_3)$ :  $\delta = 167.0$ , 165.2 (CO), 156.3 (OCONH), 104.1 (C-1,  $J_{\text{C-1-H-1}} = 158.1 \text{ Hz}$ , 101.6 (PhCH), 98.1 (C-1',  $J_{\text{C-1'-H-1'}} =$ 175.0 Hz), 82.8 (C-2), 79.1 (C-4), 76.6 (C-3), 75.1 (PhCH<sub>2</sub>), 73.3 (C-3'), 71.9 (C-4'), 70.1 (OCH<sub>2</sub>), 70.7 (C-2'), 68.7 (C-6), 68.5 (C-5'), 66.6 (PhCH<sub>2</sub>OCO), 66.4 (C-5), 40.9 (CH<sub>2</sub>NH), 29.6 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 23.2 (CH<sub>2</sub>), 17.0 (C-6'). - C<sub>53</sub>H<sub>57</sub>NO<sub>14</sub> (932.0): calcd C 68.30, H 6.16, N 1.50; found C 68.12, H 6.31, N 1.39.

5-Benzyloxycarbonylaminopentyl *O*-(2,3,4-Tri-*O*-benzyl-β-D-mannopyranosyl)-(1→4)-2-O-benzoyl-6-O-benzyl-β-D-glucopyranosyl)- $(1\rightarrow 4)$ -(2,3-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl)- $(1\rightarrow 3)$ -2-O-benzyl-4,6-*O*-benzylidene-β-D-glucopyranoside 3'',6'''-Malonate (21): Treatment of a mixture of 11 (120 mg, 0.12 mmol), 20 (116 mg, 0.12 mmol) and 3 Å molecular sieves (ca. 0.2 g) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) under argon at -70°C with NIS (28.0 mg, 0.12 mmol) and TMSOTf (11 µL, 62 µmol) for 6 h and workup as described for the preparation of 19 followed by chromatography (toluene/acetone 20:1 v/v) afforded **21** (141 mg, 64%) as a colorless foam.  $- [\alpha]_D^{20} =$ +36.1 (c = 0.8, CHCl<sub>3</sub>). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 5.65$  (dd,  $J_{2',3'} = 3.5 \text{ Hz}, 1 \text{ H}, 2'-\text{H}), 5.52 \text{ (s, 1 H, PhCH)}, 5.45 \text{ (dd, } J_{3',4'} =$ 9.8 Hz, 1 H, 3'-H), 5.22 (d,  $J_{1',2'} = 1.3$  Hz, 1 H, 1'-H), 5.14 (dd,  $J_{2'',3''} = 9.5 \text{ Hz}, 1 \text{ H}, 2''-\text{H}), 5.06 \text{ (t, s, } J_{3'',4''} = 9.3 \text{ Hz}, 1 \text{ H}, 2 \text{ H}, 3''-\text{H})$ H, PhCH<sub>2</sub>OCO), 4.72 (s, 1 H, NH), 4.71 (d,  $J_{1'',2''} = 7.8$  Hz, 1 H, 1"-H), 4.86 (d, J = -11.4 Hz, 1 H, PhCH<sub>2</sub>), 4.83 (d, J = -10.7 Hz, 1 H, PhCH<sub>2</sub>), 4.79 (d, J = -12.5 Hz, 1 H, PhCH<sub>2</sub>), 4.70 (d, J =-12.6 Hz, 1 H, PhCH<sub>2</sub>), 4.62 (d, J = -10.6 Hz, 1 H, PhCH<sub>2</sub>), 4.56 Hz $(d, J = -10.9 \text{ Hz}, 1 \text{ H}, \text{ PhCH}_2), 4.54 (d, J = -10.9 \text{ Hz}, 1 \text{ H},$ PhCH<sub>2</sub>), 4.52 (s, 1 H, PhCH<sub>2</sub>), 4.50 (dd,  $J_{6a''',6b'''} = -11.4$  Hz, 1 H, 6a'''-H), 4.48 (d, J = -12.4 Hz, 1 H, PhCH<sub>2</sub>), 4.44 (d,  $J_{1.2} =$ 7.7 Hz, 1 H, 1-H), 4.39 (d, J = -12.3 Hz, 1 H, PhCH<sub>2</sub>), 4.37–4.32 (m,  $J_{6a,6b} = -10.5$  Hz, 2 H, 6a-H, 1'''-H), 4.27-4.22 (m,  $J_{5',6'} =$ 6.2 Hz, 1 H, 5'-H), 4.05 (dd, 1 H, 6b'''-H), 4.01 (t,  $J_{4'',5''} = 9.3$  Hz, 1 H, 4"-H), 3.91-3.87 (m, 1 H, OCH<sub>2</sub>), 3.87 (t,  $J_{3,4}=9.2$  Hz, 1 H, 3-H), 3.82 (t,  $J_{4',5'}$  = 9.8 Hz, 1 H, 4'-H), 3.78 (t,  $J_{5.6b}$  = 10.5 Hz,

**FULL PAPER** G. Lemanski, T. Ziegler

1 H, 6b-H), 3.74 (t,  $J_{4''',5'''} = 9.4$  Hz, 1 H, 4'''-H), 3.67 (bd,  $J_{2''',3'''} = 3.2 \text{ Hz}, 1 \text{ H}, 2'''\text{-H}), 3.62 \text{ (t, } J_{4,5} = 9.2 \text{ Hz}, 1 \text{ H}, 4\text{-H}),$ 3.61-3.57 (m,  $J_{5''',6a'''}=7.5$  Hz,  $J_{5''',6b'''}=8.1$  Hz, 1 H, 5'''-H), 3.55-3.49 (m, 3 H, 6a"-H, 6b"-H, OCH<sub>2</sub>), 3.47-3.35 (m, 4 H, 2-H, 3'''-H, 5-H, 5"-H), 3.14-3.11 (m, 2 H, CONHCH<sub>2</sub>), 3.12 (d,  $J = -12.2 \text{ Hz}, 1 \text{ H}, \text{ COCH}_2), 3.07 \text{ (d, } J = -12.2 \text{ Hz}, 1 \text{ H},$  $COCH_2$ ), 1.66–1.35 (m, 6 H,  $CH_2$ ), 0.90 (d, 1 H, 6'-H). –  $^{13}C$ NMR (CDCl<sub>3</sub>):  $\delta = 165.6$  (COCH<sub>2</sub>), 165.1, 165.0 (1 C, 2 C, PhCO), 164.4 (COCH<sub>2</sub>), 156.3 (OCONH), 104.1 (C-1,  $J_{C-1-H-1}$  = 160.2 Hz), 103.5 (C-1''',  $J_{\text{C-1'''},\text{H-1'''}} = 154.1 \text{ Hz}$ ), 101.5 (PhCH), 100.5 (C-1",  $J_{\text{C-1''},\text{H-1''}} = 162.1 \text{ Hz}$ ), 98.3 (C-1',  $J_{\text{C-1'},\text{H-1'}} =$ 174.8 Hz), 82.6 (C-2), 82.0 (C-3'''), 78.8 (C-4), 78.5 (C-4"), 77.0 (C-3), 76.6 (2 C, C-4', C-4'''), 75.6 (C-3"), 75.1 (PhCH<sub>2</sub>), 74.9 (PhCH<sub>2</sub>), 74.6 (C-2"), 74.5 (C-5"), 73.6, 73.5 (PhCH<sub>2</sub>), 72.6 (C-3'), 72.4 (C-2"), 72.0 (PhCH<sub>2</sub>), 70.5 (C-5"), 70.4 (C-2'), 70.1 (OCH<sub>2</sub>), 68.6 (C-6), 68.4 (C-6"), 66.7 (C-5"), 66.5 (2 C, C-5, PhCH<sub>2</sub>OCO), 64.1 (C-6'''), 42.8 (COCH<sub>2</sub>), 40.8 (CH<sub>2</sub>NH), 30.0, 29.3, 23.2 (CH<sub>2</sub>), 17.2 (C-6'). - C<sub>103</sub>H<sub>105</sub>NO<sub>27</sub> (1789.0): calcd C 69.15, H 5.92, N 0.78; found C 69.09, H 6.07, N 0.69.

5-Aminopentyl O-( $\beta$ -D-Mannopyranosyl)-( $1\rightarrow 4$ )-( $\beta$ -D-glucopyranosyl)- $(1\rightarrow 4)$ - $(\alpha$ -L-rhamnopyranosyl)- $(1\rightarrow 3)$ - $\beta$ -D-glucopyranoside (22): A solution of 21 (60 mg, 33 µmol) and a catalytic amount of NaOMe in methanol (4 mL) was stirred at room temperature for 30 h, neutralized with ion exchange resin (Dowex 1X8, H<sup>+</sup> form) and concentrated. The residue was dissolved in methanol/acetic acid (1:1 v/v, 10 mL), a catalytic amount of Pd(OH)<sub>2</sub> on charcoal was added, the suspension was hydrogenized (1013 hPa H<sub>2</sub>) at room temperature for 6 d, filtered and concentrated. Chromatography of the residue with water on Biogel P2 and lyophilisation of the carbohydrate-containing fractions afforded 22 (21 mg, 87%) as a colorless foam.  $- [\alpha]_D^{20} = +73.4 (c = 1.1, H_2O). - {}^{13}C NMR$  $(D_2O)$ :  $\delta = 104.2$  (C-1''), 102.5 (C-1), 102.1 (C-1'), 101.3 (C-1'''), 83.3 (C-3), 82.3 (C-4'), 79.2 (C-4''), 77.2 (C-5), 75.1 (C-2), 75.0 (C-5"), 74.8 (C-3"), 73.9 (C-2"), 72.5 (C-3""), 72.2 (C-5""), 71.6, 71.4, 71.3 (C-2',3',2'''), 71.2 (OCH<sub>2</sub>), 70.7 (C-4'''), 69.7 (C-4), 69.1 (C-5'), 62.0 (C-6), 61.3, 61.1 (C-6",6"), 39.8 (NCH<sub>2</sub>), 28.4, 26.6, 22.2 (CH<sub>2</sub>), 17.9 (C-6').  $- C_{29}H_{53}NO_{20}$  (735.7): FAB MS (pos.); m/z: 737 (M + H<sup>+</sup>).

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